Bioactivity of *Cicuta virosa* L. var. *latisecta* Celak. (Umbelliferae: Cicutal) against Red Imported Fire Ant under Laboratory and Field Conditions

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Abstract

We have evaluated the bioactivities of compounds from *Cicuta virosa* L. var. *latisecta* Celak against red imported fire ants (*Solenopsis invicta* Buren) under field and laboratory conditions. The compounds were as follows: methanol extract; petroleum ether, chloroform, and ethyl acetate fractions from the methanol extract; and the active compound isoimperatorin, which was isolated from the chloroform-fraction. The 7 d LC50 values of the methanol extract, petroleum ether, chloroform, and ethyl acetate fractions and isoimperatorin toward micрогites were 111.20, 214.45, 40.90, 569.67, and 25.73 mg/kg, respectively. The corresponding LC50 values toward macrogites were 155.78, 308.38, 75.01, 776.75, and 42.77 mg/kg, respectively. Under field conditions, baits containing 0.2% methanol extract, 0.1% chloroform fraction, and 0.05% isoimperatorin efficiently controlled *S. invicta*, with effectiveness percentages of 95.56%, 97.78%, and 95.56%, respectively on the 30th day after bait application. Such effectiveness percentages were not significantly different from that obtained using the positive control fipronil. The present study showed that *C. virosa* L. var. *latisecta* has potential as a natural control agent for the red imported fire ants.

Introduction

The red imported fire ants, *Solenopsis invicta* Buren (Hymenoptera: Formicidae), is an exotic invasive pest ant species that has spread in many countries, including China. The imported fire ant workers sting and inject venom, which make them medically important insects (deShazo et al., 1999). Less than 3% of the human population is sensitive to the venom of fire ants, but stinging incidences have resulted in serious medical problems or even death, particularly when the victim is young, old, or has a compromised immune system (Drees et al., 2013). The fire ant has become a dominant pest in invaded areas, causing great economic loss and ecological problems.

Efforts have been made to control the red imported fire ants. Insecticide use is an important control strategy. Fipronil is a broad-spectrum pyrazole insecticide that could serve as a bait toxicant for control of ants. A 15 μg/mg granular bait caused over 80% colony mortality at 6 and 12 weeks after broadcast application in non-grazed pastures (Collins & Callcott, 1998). The activities of other insecticides, such as bifenthrin, chlorfenapyr, and thiamethoxam were also evaluated against red imported fire ants (Wiltz et al., 2010). Biological control is also an important pest management method for this species. Biological control strategies include the use of the following: *Pseudacteon* spp. (Diptera: Phoridae) parasitoids; *Kneallhazia solenopsae* (Microsporida: Tubulinosematidae) (Williams et al., 1999), which is the causative agent of fire ant disease; *Beauveria bassiana* (Balsamo) Vuillem (Bextine & Thorvilson, 2002; Oi et al., 1994); and insect viruses (Valles & Strong, 2005; Hashimoto & Valles, 2008).

Plants contain many compounds that have insecticidal activities. Many of these compounds have been used as alternatives to synthetic chemical pesticides because of their desirable attributes, such as eco-friendly nature, availability,
safety, acceptability, minimal effect on beneficial organisms, low cost, good storage, and easy application. Many plant-derived compounds have been widely used as commercial formulations, such as rotenone from the roots of *Derris trifoliata* Lour. and azadirachtin from the fruits of *Azadirachta indica* A. Juss. Plant-derived substances are globally known because they have few side effects and have insect control properties. However, information is lacking on the activities of plant-derived compounds against red imported fire ants. In our study on natural insecticides of plant origin, we obtained a methanol extract from *Cicuta virosa* L. var. *latisecta* Celak, which is a perennial herbaceous plant growing in Northeast China; this plant is a Chinese folk medicine used to treat abdominal pain (Li et al., 2009) and has potency against the red imported fire ants. We analyzed the bioactivities of the methanol extract, fractions (petroleum ether, chloroform, and ethyl acetate fractions), and an isolated active compound, namely, isoimperatorin against fire ants under laboratory and field conditions.

**Material and Methods**

**Preparation of plant extract, fractions and isoimperatorin**

The whole plant of *C. virosa* L. var. *latisecta* Celak was collected from the suburbs of Changchun, Jilin Province, China in June 2013 and was air-dried at room temperature. Methanol extract and fractions from the plant and isoimperatorin were prepared according to the author’s previously reported method (Tian et al., 2013). The powdered dry plant material was extracted with methanol at room temperature, and the methanol solution was concentrated *in vacuo*. The residue was sequentially extracted with petroleum ether, chloroform (*CHCl*$_3$), and ethyl acetate (*EtOAc*). The resulting petroleum ether, *CHCl*$_3$, and *EtOAc* solutions yielded a deep brown syrup after concentration *in vacuo*. The active compound isoimperatorin was isolated from the *CHCl*$_3$-soluble extract using the same procedure described by the author in the abovementioned reference.

**Origin and rearing of micrergate and macrergates of red imported fire ants**

*S. invicta* colonies were collected from the suburbs of Guangzhou, China and maintained in the laboratory for bioassays (Lv et al., 2006; Huang et al., 2007). The collected ants were fed with a mixture of 10% honey and live insects (*Tenebrio molitor* L.). A test tube (25 mm × 200 mm) partially filled with water and plugged with cotton was used as a water source. The ants were maintained in the laboratory at 25 ± 2 °C.

**Preparation of fire ant bait**

*C. virosa* L. var. *latisecta* methanol extract, the three abovementioned fractions, and isoimperatorin were mixed with fire ant bait for bioassays under laboratory and field conditions. The bait was prepared as described by Kafle et al. (2010, 2011) with some modifications. At first, the extract, fractions, and isoimperatorin were dissolved in acetone. Subsequently, the acetone solutions were mixed with soybean oil, shrimp shell, and corn powders. Finally, the prepared baits were placed in a ventilating cabinet for 1 h. The baits were maintained in a refrigerator (4 °C) for future use after the acetone had evaporated.

**Laboratory bioassays**

The vertical wall of a beaker with a bottom diameter of 10 cm was coated with Fluon emulsion after drying for 24 h to prevent the ants from escaping. This beaker was used as a foraging area. The artificial baits were placed 2 cm away from the inner wall of the beaker. A test tube was filled with water to approximately two-thirds full. The test tube was tightly plugged with water-saturated cotton, which was pushed 3 cm into the mouth of the test tube. The test tube was placed inside each beaker to serve as the water source for the insects.

All ants used in the toxicity tests were from the same colony. A group of 30 micrergates or macrergates was transferred into the beaker for the toxicity tests, and each test was replicated thrice. The baits, which contained the abovementioned methanol extract, fractions, or isoimperatorin at serial concentrations were placed in the beakers for the bioassay. A blank bait mixed with only acetone was used as the negative control. The mortalities were corrected by Abbott’s formula (Abbott, 1925). The average corrected mortality of three replications at each concentration was calculated 7 d later. The LC$_{50}$ value, which is the concentration that causes 50% insect mortality, was determined using probit analysis.

During the tests, mortality was recorded at an interval of 48 h for 7 d. This study was conducted under ambient temperature and relative humidity, as follows: 27 ± 1 °C and 60 ± 5 % RH on average under 14 h : 10 h light : dark photoperiod.

**Field trials**

The effectiveness of *C. virosa* L. var. *latisecta* baits containing methanol extract, chloroform fraction, and isoimperatorin on the inactivation of *S. invicta* mounds were evaluated under field conditions. The field studies were conducted in the Zengcheng Farm of South China Agricultural University. The experiment was designed in a single factor random block. Fifteen mounds were used for each bait and for control treatments. Each treatment was replicated thrice. The baits were sprinkled around a 50 cm radius from the mounds’ center. The commercial insecticide fipronil (purity 96%, Tuo Qiu Agricultural Chemical Ltd. Co., Jiangshu Province, China) was used as the positive control. The blank bait mixed with only acetone was used as the negative control. The number of active mounds in the field was determined on the 1st, 5th, 10th, 20th, and 30th days after
treatment. A mound was designated as active when at least 20 adult workers exited the excavated soil when probed with a metal rod (5 mm in diameter) (Kafle et al., 2011). The control effectiveness was calculated using the following equation:

\[
\text{control effectiveness} = \left(\frac{S_{\text{control}} - S_{\text{treatment}}}{100 - S_{\text{control}}}\right) \times 100,
\]

where

\[S = \frac{\text{number of active mounds before treatment} - \text{number of active mounds after treatment}}{\text{number of active mounds before treatment}} \times 100.
\]

**Statistical analyses**

The results were expressed as mean ± standard error. The treatment means were transformed into arcsine square-root values for analysis of variance (ANOVA). The means were compared and separated by Scheffe’s test. \(P < 0.05\) was the level of statistical significance.

**Results and Discussion**

**Laboratory bioassays**

*C. virosa* L. var. *latisecta* methanol extract and fractions (petroleum ether, CHCl₃, and EtOAc) and isoimperatorin showed insecticidal activities against micrergates of red imported fire ants (Fig 1). The insecticidal activities were slow to show, but the corrected percentage mortalities of treated micrergates increased steadily with increasing treatment time. Seven days after the treatment, the highest mortality (93.33%) occurred among the micrergates treated with 600 mg/kg methanol extract. The micrergate mortalities under the other treatments were in the following descending order: 200 mg/kg chloroform fraction (90.00%), 40 mg/kg isoimperatorin (70.00%), 200 mg/kg petroleum ether fraction (45.00%), 200 mg/kg ethyl acetate fraction (28.33%), and the control (3.33%). The 7 d \(L_{C50}\) values of the methanol extract and fractions (petroleum ether, CHCl₃, and EtOAc) and isoimperatorin for the micrergates were 111.20, 214.45, 40.90, 569.67, and 25.73 mg/kg, respectively (Table 1).

![Fig 1. Mortalities of Cicuta virosa L. var. latisecta methanol extract, fractions and isoimperatorin against micrergates of Solenopsis invicta under laboratory conditions.](image1)

![Fig 2. Mortalities of Cicuta virosa L. var. latisecta methanol extract, fractions and isoimperatorin against macrergates of Solenopsis invicta under laboratory conditions.](image2)

For the macrergates, similar trends were observed (Fig 2). The 600 mg/kg methanol extract resulted in the highest mortality (80.00%) at 7 d after treatment. The mortalities induced by the other treatments were in the following decreasing order: 200 mg/kg chloroform fraction (71.67%), 40 mg/kg isoimperatorin (53.33%), 200 mg/kg petroleum ether fraction (35.00%), 200 mg/kg ethyl acetate fraction (23.33%), and the control (1.67%). The 7 d \(L_{C50}\) values of the methanol extract and fractions (petroleum ether, CHCl₃, and EtOAc) and isoimperatorin for macrergates were 155.78, 308.38, 75.01, 776.75, and 42.77 mg/kg, respectively (Table 2).

**Table 1.** \(L_{C50}\) of Cicuta virosa L. var. latisecta methanol extract, petroleum ether-, chloroform- and ethyl acetate fractions, and isoimperatorin against micrergates of Solenopsis invicta.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Regression equation</th>
<th>(L_{C50}) (mg/kg)</th>
<th>95% Fiducial limit</th>
<th>Correlation coefficients</th>
</tr>
</thead>
<tbody>
<tr>
<td>Methanol extract</td>
<td>(Y = 1.3499 + 1.7839x)</td>
<td>111.20</td>
<td>89.76 ~ 137.76</td>
<td>0.9917</td>
</tr>
<tr>
<td>Petroleum ether fraction</td>
<td>(Y = 0.8122 + 1.7963x)</td>
<td>214.45</td>
<td>175.07 ~ 262.69</td>
<td>0.9868</td>
</tr>
<tr>
<td>Chloroform fraction</td>
<td>(Y = 2.0932 + 1.8035x)</td>
<td>40.90</td>
<td>33.26 ~ 50.29</td>
<td>0.9907</td>
</tr>
<tr>
<td>Ethyl acetate fraction</td>
<td>(Y = -0.0535 + 1.8339x)</td>
<td>569.67</td>
<td>466.83 ~ 695.18</td>
<td>0.9966</td>
</tr>
<tr>
<td>Isoimperatorin</td>
<td>(Y = 2.4614 + 1.7999x)</td>
<td>25.73</td>
<td>21.03 ~ 31.48</td>
<td>0.9898</td>
</tr>
</tbody>
</table>
Table 2. LC50 of Cicuta virosa L. var. latisecta methanol extract, petroleum ether-, chloroform- and ethyl acetate fractions, and isoimperatorin against macrergates of Solenopsis invicta.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Regression equation</th>
<th>LC50 (mg/kg)</th>
<th>95% Fiducial limit</th>
<th>Correlation coefficients</th>
</tr>
</thead>
<tbody>
<tr>
<td>Methanol extract</td>
<td>$Y = 1.2080 + 1.7295x$</td>
<td>155.78</td>
<td>126.60 ~ 191.67</td>
<td>0.9946</td>
</tr>
<tr>
<td>Petroleum ether fraction</td>
<td>$Y = 0.0045 + 2.0070x$</td>
<td>308.38</td>
<td>256.17 ~ 371.23</td>
<td>0.9899</td>
</tr>
<tr>
<td>Chloroform fraction</td>
<td>$Y = 1.9671 + 1.6174x$</td>
<td>75.01</td>
<td>58.85 ~ 95.61</td>
<td>0.9938</td>
</tr>
<tr>
<td>Ethyl acetate fraction</td>
<td>$Y = -0.5601 + 1.9237x$</td>
<td>776.75</td>
<td>636.97 ~ 947.18</td>
<td>0.9902</td>
</tr>
<tr>
<td>Isoimperatorin</td>
<td>$Y = 1.9383 + 1.8771x$</td>
<td>42.77</td>
<td>35.08 ~ 52.13</td>
<td>0.9907</td>
</tr>
</tbody>
</table>

Field trials

C. virosa L. var. latisecta methanol extract, chloroform fraction, and isoimperatorin could control S. invicta, but these botanical insecticides were slower to act than fipronil (Table 3). On the 5th day after bait application, the control effectiveness of baits containing 0.2% methanol extract, 0.1% chloroform fraction, and 0.05% isoimperatorin were 33.33%, 31.11%, and 26.67%, respectively. These results were significantly lower than the positive control fipronil (66.67%) ($F = 14.297$, $P = 0.0039$). On the 10th day, the control effectiveness value of 0.01% fipronil-based bait was still significantly higher than the three baits ($F = 11.407$, $P = 0.0068$). However, on the 30th day, the control effectiveness of the baits containing methanol extract, chloroform fraction, and isoimperatorin increased to 95.56%, 97.78%, and 95.56%, respectively, and such percentages were not significantly different from the control effectiveness percentage of fipronil bait (100%) ($F = 2.2$, $P = 0.1889$). However, the control value of 0.01% fipronil bait treatment remained the highest throughout the 30 d period after bait application.

In the present study, the bioactivity of the insecticidal plant (C. virosa var. latisecta) against the red imported fire ant was investigated. The crude methanol extract and fractions (petroleum ether, CHCl3, and EtOAc) and the isolated compound isoimperatorin were verified to be active against micrergates and macrergates of the ants under both laboratory and field conditions. C. virosa L. var. latisecta compounds are safe for use as control agents of fire ants. C. virosa L. var. latisecta is used in Northeast China to control aphids on vegetables. Furthermore, the isolated active compound, isoimperatorin, is also a main active constituent in other safe-for-use traditional Chinese medicinal plants, such as Notopterygium forbesii Boiss (Zhou et al., 2007). C. virosa L. var. latisecta has potential as a source of natural insecticides for the control of the red imported fire ants.

Table 3. Controlling effectiveness of Cicuta virosa L. var. latisecta methanol extract-, chloroform fraction-, isoimperatorin- and fipronil-based baits on the inactivation of Solenopsis invicta mounds under field conditions.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Controlling effectiveness (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1d after treatment</td>
</tr>
<tr>
<td>0.2% Methanol extract-based bait</td>
<td>0.00 ± 0.00 b</td>
</tr>
<tr>
<td>0.1% Chloroform fraction-based bait</td>
<td>0.00 ± 0.00 b</td>
</tr>
<tr>
<td>0.05% Isoimperatorin-based bait</td>
<td>0.00 ± 0.00 b</td>
</tr>
<tr>
<td>0.01% fipronil-based bait</td>
<td>15.56 ± 2.22 a</td>
</tr>
</tbody>
</table>

References


