



SHORT NOTE

A Short Note on an Artisanal Incubator for Fermentation of *Apis mellifera* Artificial Diets

JPLM PAIVA¹, C GAMA¹, PM DRAGO², C BARBIERI², MM MORAIS³

1 - Universidade Federal de São Paulo, Instituto de Ciência e Tecnologia, São José dos Campos-SP, Brazil

2 - Universidade de São Paulo, Escola de Artes, Ciências e Humanidades, São Paulo-SP, Brazil

3 - Universidade Federal de São Paulo, Departamento de Ecologia e Evolução, Diadema-SP, Brazil

Article History

Edited by

Evandro N. Silva, UEFS, Brazil

Received 25 November 2019

Initial acceptance 05 December 2019

Final acceptance 05 December 2019

Publication date 30 December 2019

Keywords

Fermentative process, supplementary diet, honey bees, apiculture.

Corresponding author

Juliana Pereira Lisboa M. Paiva

Instituto de Ciência e Tecnologia

Universidade Federal de São Paulo – Unifesp

Rua Talim, 330, Vila Nair - CEP: 12231-280

São José dos Campos, São Paulo, Brasil.

E-Mail: jplbiologia@gmail.com

Abstract

Considering the importance of offering food supplementation to the swarms during dearth periods, we developed in this project an artisanal incubator for fermentation of supplementary protein diets for *Apis mellifera* bees, obtaining a fresh, nutritious and palatable product, made on the property, thus facilitating access to the beekeeper to this resource.

With the growing need for food supplementation of swarms, especially in periods of reduced food supply in the wild, it is important to provide technological resources directly to producers, so that they can produce on their property the food that bees need. Research reports the importance of nutritional supplementation, especially protein, when pollen sources in nature suffer significant reductions, colonies reduce their productivity due to their weakening (Saffari et al., 2010; Morais et al., 2013; Paiva et al., 2019). However, the fermentation of this food becomes a fundamental factor in the animal's acceptance and consumption, since this microbiological process makes the food closer to the beebread: more palatable and attractive (Ellis & Hayes Jr, 2009; Li et al., 2012).

For the fermentation process to occur satisfactorily, the control of factors such as temperature guarantee the performance of the fermenting microorganisms (especially

of the genus *Lactobacillus* and *Pediococcus*) that find in the range of 27° to 32°C the ideal point for the fermentation of sugars (sucrose) present in the food, converting them into lactic acid (Kristensen et al., 2010; Kung et al., 2011). This process reduces the pH of the food to 3.8, conserving its nutrients and preventing the attack of spoilage organisms, especially bacteria of the genus *Clostridium*, capable of consuming the amino acids of the food, reducing its palatability and nutritional value (Shao et al., 2002; Schröder et al., 2013; Strauber et al., 2016). Thus, the food preserves its desirable characteristics, being consumed and used by the swarms, strengthening them and improving their productive performance, even in periods of reduced floral resources (Paiva et al., 2016; Paiva et al., 2019).

The availability of a simple and inexpensive equipment such as incubator allows the producer to ferment the amount of food needed and store it in perfect condition until it is used by



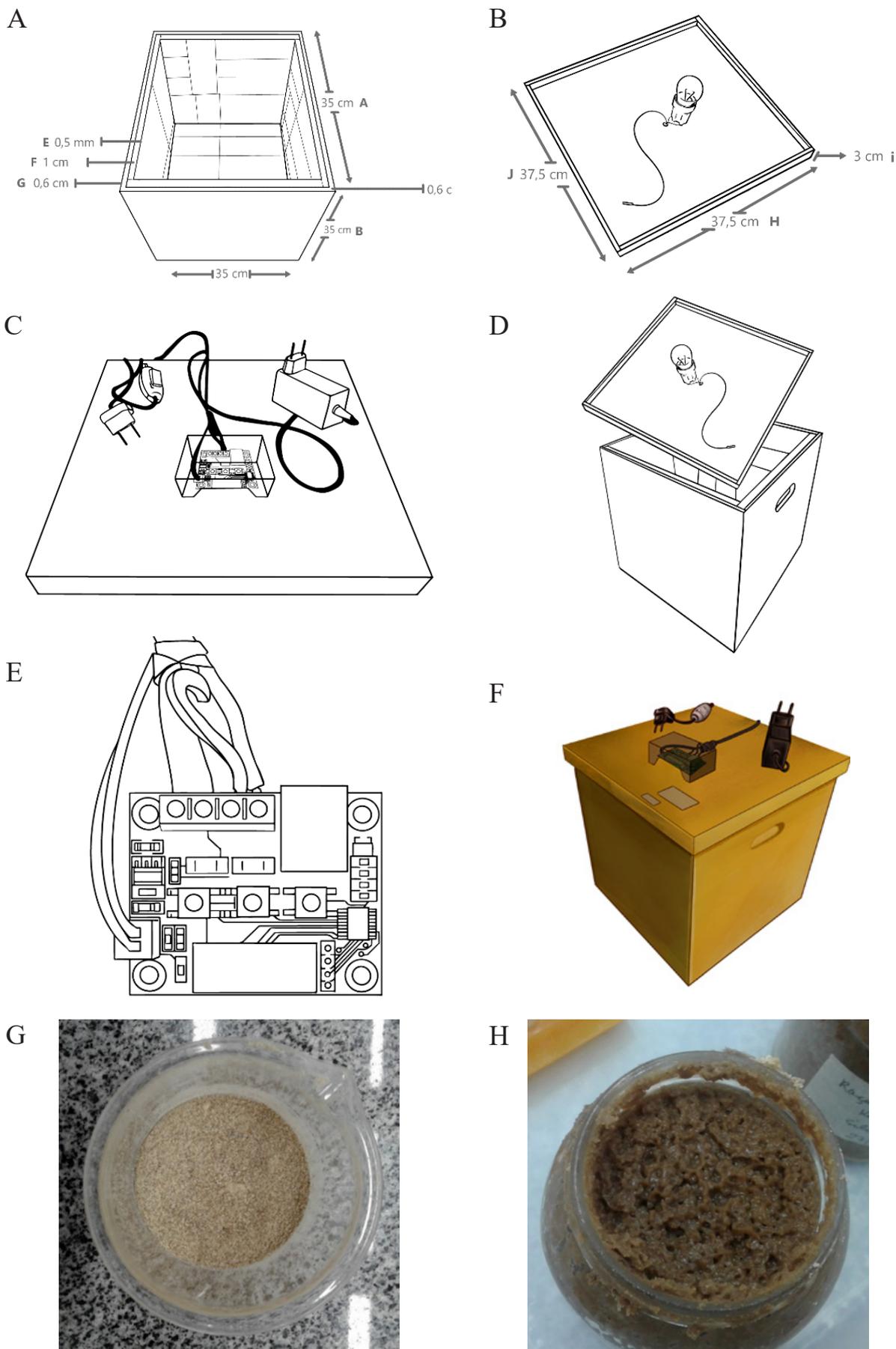


Fig 1. Details of the constitution of the artisanal incubator: A. Styrofoam and polyethylene lined box; B. cover with socket and lamp; C. thermostat and socket; D. Inner face of box and lid; E. digital thermostat; F. ready incubator; G. diet before fermentation and H. fermented diet.

the swarms. It can prevent nutrient loss and deterioration and offer to the bees a beneficial, fresh, efficient food as it would avoid stocking the product for very long periods, causing diet rejection (Carroll et al., 2017). According to this, the objective of this work was to create a facilitating artisanal incubator for beekeepers and stingless beekeepers interested in using fermented feed in their bee pasture, enabling the fermentation of bees feed and maintaining the temperature between 27° and 28°, reaching 30°C. For this purpose, materials such as a styrofoam box (40x40x30cm) were used for the prototype. It was later replaced by a wooden box (35x35x35 cm) with a styrofoam coated lid and thermally insulated with carton packs lined with polyethylene and aluminum layers (used in milk or juice boxes), digital thermostat, porcelain socket; 15w light bulb; parallel wire; switch and socket (Figure 1). System operation occurs through the digital thermostat, which has a minimum and maximum temperature setting display with a relay on/off system, as well as a thermal sensor, which reads the ambient temperature, showing it on the display.

With set temperatures from 28°C minimum and 30°C maximum, the thermostat allows current to flow to the lamp when the sensor registers temperatures below the maximum. Thus, the lamp lights generating heat inside the box. When the sensor records a maximum temperature of 30°C, the system will no longer allow current to pass, so turning off the lamp.

After the incubator confection, four capped glass jars containing 100g of soybean meal, corn, sugarcane yeast, egg powder and vitamin-mineral premix feeds were inserted (Paiva et al., 2019). 40 ml of 50% sucrose solution and 0.5 mg of microbial inoculum (*Lactobacillus plantarum* and *Pediococcus sp*) were added to each flask to form a paste. These flasks were kept for 7 days, 14 and 21 days at 30°C, with daily process monitoring until completion. The pH of the rations was measured on the 1st, on the 7th day, on the 14th and 21st day of the experiment, for follow-up.

After seven days, we observed that the rations fermented satisfactorily, presenting the formation of bubbles in the mass (due to the release of gases from microbial activity) and pH change (initial: 4.76; 7th day: 3.62; 14th day: 3.67 and 21st day, 3.77), the reduction of which is evidence of acid production by microorganisms from the fermentation of sugars. After the process was completed, 4g of feed were offered to 60 cage-confined worker bees, with 7 repetitions, for consumption of the fermented feed, which was well accepted by animals (Paiva et al., 2019).

Considering the context, we can conclude that the use of affordable and low cost materials has enabled the production of an efficient artisanal incubator, allowing the beekeeper to ferment and conserve supplementary protein food for bees, especially during periods of greatest pollen deficiency in nature. With this resource, the beekeeper can have an effective food available, minimizing losses and helping the swarms to maintain the beekeeping production.

Acknowledgments

The authors would like to thank Ms. Joanize Paiva for comments on the use of the English language.

References

- Carroll, M.J., Brown, N., Goodall, C., Downs, A.M., Sheenan, T.H. & Anderson, K.E. (2017). Honey bees preferentially consume freshly-stored pollen. *PloS One*, 12(4), e0175933. doi: 10.1371/journal.pone.0175933
- Ellis, A.M. & Hayes Jr, G.W. (2009). An evaluation of fresh versus fermented diets for honey bees (*Apis mellifera*). *Journal of Apicultural Research*, 48: 215-216. doi: 10.3896/IBRA.1.48.3.11
- Kung, L. & Ranjit, N.K. (2011). The Effect of *Lactobacillus buchneri* and Other Additives on the Fermentation and Aerobic Stability of Barley Silage. *Journal of Dairy Science*, 84: 1149-1155. doi: 10.3168/jds.S0022-0302(01)74575-4
- Kristensen, N.B., Sloth, K.H., Højberg, O., Spliid, N.H., Jensen, C. & Thøgersen, R. (2010). Effects of microbial inoculants on corn silage fermentation, microbial contents, aerobic stability, and milk production under field conditions. *Dairy Science*, 93: 3764-3774. doi: 10.3168/jds.2010-3136
- Li, C., Xu, B., Wang, Y., Feng, Q. & Yang, W. (2012). Effects of dietary crude protein levels on development, antioxidant status, and total midgut protease activity of honey bee (*Apis mellifera ligustica*). *Apidologie*, 43: 576-586. doi: 10.1007/s13592-012-0126-0
- Morais, M.M., Turcatto, A.P., Pereira, R.A., Franco, T.M., Guidugli-Lazzarini, K.R., Gonçalves, L.S., De Almeida, J.M. Ellis, J.D. & De Jong, D. (2013). Protein levels and colony development of Africanized and European honey bees fed natural and artificial diets. *Genetics and Molecular Research*, 12: 6915-6922.
- Paiva, J.P.L.M., Paiva, H.M., Esposito, E. & Morais, M.M. (2016). “On the Effects of Artificial Feeding on Bee Colony Dynamics: A Mathematical Model”. *PloS One*, 11 (11): e0167054. doi: 10.1371/journal.pone.0167054
- Paiva, J.P.L.M., Esposito, E., Souza, G.I.H., Franco, T.M. & Morais, M.M. (2019). Effects of ensiling on the quality of protein supplements for honey bees *Apis mellifera*. *Apidologie*, 50: 414-424, doi: 10.1007/s13592-019-00661-4
- Saffari, A., Kevan, P.G. & Atkinson, J.L. (2010). Palatability and consumption of patty-formulated pollen and pollen substitutes and their effects on honeybee colony performance. *Journal of Apicultural Science*, 54: 63-69.
- Shao, T., Ohba, N., Shimojo, M. & Masuda, Y. (2002). “Dynamics of Early Fermentation of Italian Ryegrass (*Lolium multiflorum* Lam.) Silage. *Asian-Australian Journal of Animal Science*, 15: 1606-1610. doi: 10.5713/ajas.2002.1606

Schröder, J.J., Visser, W., Assinck, F.B.T. & Velthof, G.L. (2013). Effects of short-term nitrogen supply from livestock manures and cover crops on silage maize production and nitrate leaching. *Soil Use and Management*, 29: 151-160. doi: 10.1111/sum.12027

Strauber, H., Lucas, R. & Kleinsteuber, S. (2016). Metabolic and microbial community dynamics during the anaerobic digestion of maize silage in a two-phase process. *Applied Microbiology and Biotechnology*, 100: 479-491. doi: 10.1007/s00253-015-6996-0

