



## SHORT NOTE

### Karyotype Description of *Cephalotrigona femorata* Smith (Hymenoptera: Apidae) and the C-banding Pattern as a Specific Marker for *Cephalotrigona*

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#### Abstract

*Cephalotrigona femorata* (Smith, 1854) was submitted to cytogenetic techniques to study and describe its karyotype. Conventional staining allowed the counting ( $2n=34$ ) and observation of chromosome morphology. The amount and distribution of heterochromatin in this species was different from *Cephalotrigona capitata* (Smith, 1854), another species of the genus already analyzed. Our results indicate that heterochromatin is a potential marker for the genus, at least for the species found in Brazil. This region was marked by DAPI, revealing a high content of A:T. The  $CMA_3$  marked two pairs, and it seems to be polymorphic in one pair.

Cytogenetic analyses allow for morphological and quantitative characterization of the chromosomes of a determined species, which may contribute to the understanding of phylogeny and improvements in the taxonomy of some groups of Hymenoptera (Rocha et al., 2003). A very peculiar taxon within Hymenoptera is the tribe Meliponini, the stingless bees. These bees play an important role in economic and ecological processes of tropical regions because they are important pollinators of native plants (Heard, 1999; Michener, 2000). In Brazil two species of the genus *Cephalotrigona* Schwarz, 1940 are found among the five species described taxonomically, *Cephalotrigona capitata* and *C. femorata* (Moure, 2011). *Cephalotrigona capitata* is the only species of the genus for which the karyotype has already been described, and  $2n = 34$  chromosomes were observed (Rocha et al., 2003). The present study evaluates the species *C. femorata* in order to characterize it cytogenetically and thus to obtain more knowledge on stingless bees.

Post-defecant larvae of four colonies of *C. femorata*, were obtained in the municipality of Urbano Santos, Maranhão, Brazil ( $3^{\circ}12'29''S$ ;  $43^{\circ}24'18''W$ ), as part of a project to rescue wildlife in the area of deforestation. Larvae ( $n=40$ ) were collected and processed according to the method described by Imai et al. (1988) to acquire the mitotic metaphase chromosomes. Conventional staining was performed using diluted Giemsa in Sørensen buffer (0.006 M, pH 6.8) at the concentration of 4% for 20 minutes. For C-banding the BSG (Barium hydroxide/Saline/Giemsa) method was used according to Sumner (1972). Fluorochrome staining ( $CMA_3/DA/DAPI$ ) was performed according to the protocol proposed by Schweizer (1980). Ten slides from each hive were analyzed along with an average of 10 metaphases per slide. The material was observed under an Olympus BX60 microscope and images were used for assembly of the karyotypes utilizing the image analysis program Image-Pro Plus™ (version 6.3, Media Cybernetics®, 2009). Karyo-



types were organized as reported by Imai (1991), taking into consideration heterochromatin distribution.

The conventional staining technique showed that *C. femorata* presents a diploid number of  $2n = 34$  in females (Fig. 1A and B). This data is similar to that of the other species of the genus, *C. capitata*. The C-band technique permitted observation of heterochromatic arms on all chromosomes (Fig. 1B). Often these portions represent regions larger than the euchromatic arms. Two pairs also showed heterochromatin in the pericentromeric region (Fig. 1B). The observed heterochromatin pattern differed significantly from *C. capitata* where only 8 of the 17 pairs had a heterochromatin arm. According to the classification proposed by Imai (1991), *C. femorata* had a karyotypic formula of  $2k=30A^M+4A^{Mc}$  while for *C. capitata* this was  $2K=18A+16A^M$ , this variation can therefore be used to distinguish the two species since they are morphologically similar.

The heterochromatic regions were positively stained by DAPI revealing a high content of A:T in heterochromatin similar to other Meliponini (Fig. 1C). The fluorochrome CMA<sub>3</sub> showed four markings. In one of the pairs this staining showed a large polymorphism of size (Fig. 1C) in one homolog. These polymorphisms are common in bees and have been well characterized for other stingless bee species such as *Oxytrigona flaveola* (Krinski et al., 2010). As previously observed in other stingless bees, these GC-rich regions

may be related to the nucleolus organizer regions (Brito et al., 1997; Maffei et al., 2001; Rocha, 2002; Brito-Ribon et al., 2005; Lopes et al., 2011).

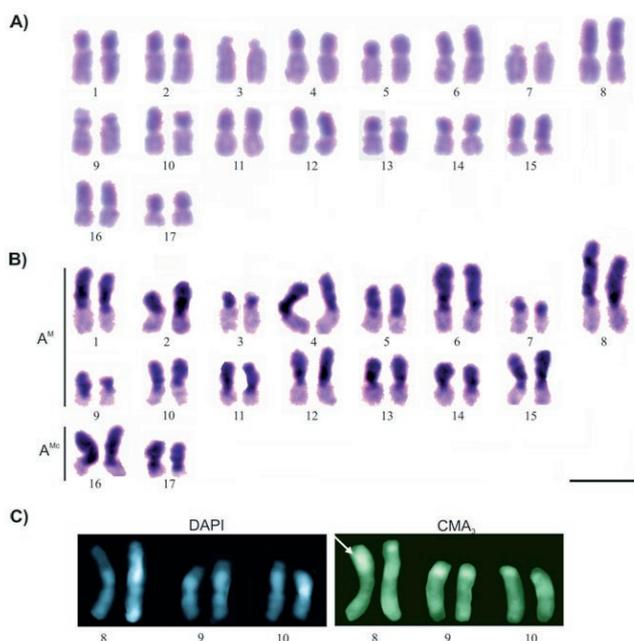
Thus, although the number of chromosomes in general is constant in the genus, other characteristics such as the amount and distribution of heterochromatin vary among species and may be used as an important tool that may aid in the taxonomy and conservation of this species.

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**Figure 1.** Karyotype of a *Cephalotrigona femorata* female. A. Conventional staining. B: C-banding. C: Fluorochromes DAPI and CMA<sub>3</sub>. Polymorphic pair indicated by arrow. Scale bar: 5  $\mu$ m.

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